Design of Clinical Trials for Development of Molecularly Targeted Drugs

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Research Areas

Clinical trials, Drug Discovery, Molecular Cancer Diagnosis, Biomedical Imaging, Computational and Systems Biology, and Biostatistical Research

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Sample Size Calculation

Mathematics And Oncology

- The Norton-Simon Hypothesis
- The Norton-Simon Hypothesis and Breast Cancer Mortality in National Randomized Trial

BRB Annual Report 2005

Position Available

Post-doctoral fellow positions available

Software Download

- Accelerated Titration Design Software
- Optimal Two-Stage Phase II Design Software
Objectives of Phase I Trials

- Develop dose/schedule
- Determine whether the drug inhibits the targeted pathway
Dose/Schedule

• Ideal is to have a drug and target so specific for cancer cells that the drug can be delivered repeatedly at doses that completely shut down the de-regulated pathway without toxicity to normal cells

• Because most current targets are not specific to cancer cells, most targeted drugs are toxic
Dose/Schedule

- Few examples of drugs whose effectiveness at inhibiting target decreases with dose after maximum
- Optimizing dose for maximum inhibition of target is difficult due to assay variability and need for tumor biopsies
- Titrating dose to achieve a pre-specified plasma concentration at which target is inhibited in pre-clinical systems is more feasible
Dose/Schedule

• Determining dose just below MTD which can be delivered repeatedly is often the most practical approach

• Accrue an additional cohort of patients at that selected dose to determine whether the target is inhibited
Components of a Phase I Design

- Starting dose
- Dose increments
- Patients per cohort
- Decision rules for dose assignment
- Intra-patient dose modification rules
- Stopping rule
- Method of analysis
Conventional Phase I Designs

• Starting dose $1/10^{th} \text{LD}_{10}$ in most sensitive species

• Modified Fibonacci dose steps
  – 100%, 67%, 50%, 40%, 33%, 33%, …

• Cohorts of 3-6 new patients per dose level

• Define MTD as highest dose with <33% DLT

• Use first course information only

• Use DLT vs non-DLT dichotomy

• No intra-patient dose escalation
Enter 3 patients

- 0/3
- 1/3
- >1/3

Enter 3 patients at same dose

- 1/6
- >1/6

Escalate dose for next cohort

MTD exceeded
Limitations of Conventional Phase I Trial Designs

- Many patients may be treated at very low doses
- Trial may take a long time to complete
- Limited information yield
Accelerated titration designs for phase I clinical trials in oncology

### Cohort Escalation Designs

<table>
<thead>
<tr>
<th></th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Cohorts of 3 new patients per dose level with 40% dose increments. If 1 of 3 experience DLT in first course, expand to cohort of 6</td>
</tr>
<tr>
<td>2</td>
<td>Cohorts of 1 new patient per dose level. When first instance of first course DLT or second instance of first course grade 2 toxicity is observed, revert to design 1.</td>
</tr>
<tr>
<td>3</td>
<td>Same as design 2 except that double dose steps are used during accelerated stage.</td>
</tr>
<tr>
<td>4</td>
<td>Cohorts of 1 new patient per dose level and double dose steps. When first instance of any course DLT or second instance of any course grade 2 toxicity is observed, revert to design 1.</td>
</tr>
</tbody>
</table>
## Within Patient Escalation Options

<table>
<thead>
<tr>
<th></th>
<th>No within patient dose escalation</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>B</strong></td>
<td>Escalate if grade 0-1 toxicity at previous course.</td>
</tr>
<tr>
<td></td>
<td>De-escalate if grade 3+ toxicity at previous course.</td>
</tr>
<tr>
<td></td>
<td>Do not assign dose at which 2 previous pts have experienced 3+ toxicity at that course or earlier.</td>
</tr>
</tbody>
</table>
Testing the 8 Designs

We fit the model to 20 phase I trials, relating to:

- Flavone acetic acid (5)
- Piroxantrone (2)
- Chloroquinoxaline sulfonamide (2)
- Pyrazoloacridine (1)
- Cyclopentenylcytosine (1)
- Fostriecin (2)
- 9-Aminocamptothecin (2)
- Penclomadine (2)

For each trial, we performed 1000 simulations for each of the 8 designs, using the model.

We compiled the results to compare the performances of the 8 designs.
Model Relating Toxicity to Dose

\[ Y_{ij} = \log\left(d_{ij} + \alpha D_{ij}\right) + \beta_i + \varepsilon_{ij} \]

\( d_{ij} \) = dose for i'th patient in course j
\( D_{ij} \) = cumulative dose up to course j for patient i
\( \alpha \) = cumulative toxicity parameter
\( \beta_i \) = patient specific effect
\( \varepsilon_{ij} \) = course specific random variation
Model Relating Toxicity to Dose

\[ Y_{ij} = \log (d_{ij} + \alpha D_{ij}) + \beta_i + \epsilon_{ij} \]

- \( Y_{ij} < K_1 \) grade 0-1 toxicity
- \( K_1 < Y_{ij} < K_2 \) grade 2 toxicity
- \( K_2 < Y_{ij} < K_3 \) grade 3 toxicity
- \( Y_{ij} > K_3 \) grade 4 toxicity
### Estimates of Parameters for 20 Clinical Trials

<table>
<thead>
<tr>
<th>Drug</th>
<th>( \alpha )</th>
<th>((K_1 - \ln d_0)/\ln 1.4)</th>
<th>((K_2 - K_1)/\ln 1.4)</th>
<th>((K_3 - K_2)/\ln 1.4)</th>
<th>(\sigma_\beta)</th>
<th>(\sigma_\epsilon)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Flavone acetic acid</td>
<td>0</td>
<td>16.2</td>
<td>6.9</td>
<td>35 no grade 4</td>
<td>0.26</td>
<td>1.9</td>
</tr>
<tr>
<td>Flavone acetic acid</td>
<td>0</td>
<td>16.1</td>
<td>8.4</td>
<td>29 no grade 4</td>
<td>2.9</td>
<td>0.85</td>
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<tr>
<td>Flavone acetic acid</td>
<td>0</td>
<td>4.4</td>
<td>2.4</td>
<td>0.95</td>
<td>0.47</td>
<td>0.59</td>
</tr>
<tr>
<td>Flavone acetic acid</td>
<td>0.24</td>
<td>8.0</td>
<td>2.9</td>
<td>2.2</td>
<td>0</td>
<td>0.83</td>
</tr>
<tr>
<td>Flavone acetic acid</td>
<td>0</td>
<td>18.5</td>
<td>6.4</td>
<td>20 no grade 4</td>
<td>0.006</td>
<td>2.8</td>
</tr>
<tr>
<td>Piroxantrone</td>
<td>0.08</td>
<td>8.4</td>
<td>2.7</td>
<td>2.3</td>
<td>1.03</td>
<td>0.42</td>
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<tr>
<td>Piroxantrone</td>
<td>0</td>
<td>16.4</td>
<td>13.3 no grade 3+</td>
<td>9.5 no grade 3+</td>
<td>0</td>
<td>1.8</td>
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<tr>
<td>Chloroquinoxaline</td>
<td>0.04</td>
<td>17.3</td>
<td>2.6</td>
<td>1.6</td>
<td>0.88</td>
<td>0.87</td>
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<tr>
<td>Chloroquinoxaline</td>
<td>0</td>
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<td>4.6</td>
<td>2.9</td>
<td>0.62</td>
<td>0.90</td>
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<tr>
<td>Pyrazine diazohydroxide</td>
<td>2.5</td>
<td>12.0</td>
<td>4.1</td>
<td>5.8</td>
<td>1.3</td>
<td>1.5</td>
</tr>
<tr>
<td>Pyrazine</td>
<td>0.24</td>
<td>6.6</td>
<td>1.3</td>
<td>0.53</td>
<td>0.002</td>
<td>0.65</td>
</tr>
<tr>
<td>Pyrazine</td>
<td>0.02</td>
<td>4.6</td>
<td>0.53</td>
<td>0.56</td>
<td>0.001</td>
<td>0.18</td>
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<tr>
<td>Pyrazoloacrine</td>
<td>0.04</td>
<td>8.9</td>
<td>1.0</td>
<td>1.3</td>
<td>0.24</td>
<td>0.32</td>
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<tr>
<td>Cyclopentomyl</td>
<td>0</td>
<td>4.4</td>
<td>0.83</td>
<td>0.18</td>
<td>0.21</td>
<td>0.27</td>
</tr>
<tr>
<td>Fostriecin</td>
<td>0.04</td>
<td>3.5</td>
<td>3.6</td>
<td>4.5</td>
<td>1.06</td>
<td>0.54</td>
</tr>
<tr>
<td>Fostriecin</td>
<td>0</td>
<td>6.3</td>
<td>7.2</td>
<td>18 no grade 4</td>
<td>0.58</td>
<td>1.6</td>
</tr>
<tr>
<td>9-AC</td>
<td>0</td>
<td>6.4</td>
<td>0.48</td>
<td>0.39</td>
<td>0.24</td>
<td>0.11</td>
</tr>
<tr>
<td>9-AC</td>
<td>0</td>
<td>6.0</td>
<td>0.51</td>
<td>1.1</td>
<td>0.35</td>
<td>0.27</td>
</tr>
<tr>
<td>Penclomadine</td>
<td>0.05</td>
<td>6.0</td>
<td>3.7</td>
<td>15 no grade 4</td>
<td>0.68</td>
<td>0.81</td>
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<tr>
<td>Penclomadine</td>
<td>0</td>
<td>5.8</td>
<td>2.0</td>
<td>17 no grade 4</td>
<td>0.43</td>
<td>0.53</td>
</tr>
</tbody>
</table>
Patients and Cohorts Distribution for 8 Designs

Average: 20 Trials

Total patients

Cohorts

Design 1A
Design 1B
Design 2A
Design 2B
Design 3A
Design 3B
Design 4A
Design 4B
Toxicity Distribution for 8 Designs

Percent of patients

- No toxicity
- Mild toxicity
- DLT
- Unaccept. toxicity

Design 1A
Design 1B
Design 2A
Design 2B
Design 3A
Design 3B
Design 4A
Design 4B
Toxicity Distribution for 8 Designs

- No toxicity
- Mild toxicity
- DLT
- Unaccept. toxicity

Design 1A
Design 1B
Design 2A
Design 2B
Design 3A
Design 3B
Design 4A
Design 4B
Accelerated Titration Designs

• Reduces patient under-treatment
  – 1 patient per dose level
  – intra-patient dose escalation

• Reduces number of patients
  – 1 patient per dose level
  – dose doubling until toxicity

• Improves information yield
  – cumulative toxicity
  – inter-patient variability
Software Available

• S+ function to fit model to phase I data
  – Point and interval estimates of parameters
  – Graphical representation of dose/response

• Excel spreadsheet and macro for quality control of dose level assignment and maintenance of dose/toxicity data

• Available at http://linus.nci.nih.gov/~brb
Phase I Designs Using Biological Endpoint

• Dose giving plasma concentration at which target is inhibited in animal model
  • Titration within patients

• Optimizing dose for target inhibition in tumor or surrogate tissue
Korn et al Phase I Design for Finding Biologically Active Dose

Buolamwini & Adjei, Novel Anticancer Drug Protocols, Humana 2003

• Treat one patient per dose level until one biological response is seen
• After the first response, treat cohorts of 3-6 patients per dose
  – With 0-1 responses in 3 patients, escalate dose for next cohort
  – With 2-3 responses in 3 patients, expand cohort to 6 patients
  – With 5-6 responses, end
  – With <5 responses, escalate dose for next cohort
## Phase I Determination of Minimum Biologically Active Dose

<table>
<thead>
<tr>
<th>Probability of Biological Response</th>
<th>Number of Patients Treated at Dose</th>
<th>Probability of No Biological Responses</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.20</td>
<td>11</td>
<td>0.09</td>
</tr>
<tr>
<td>0.25</td>
<td>9</td>
<td>0.08</td>
</tr>
<tr>
<td>0.30</td>
<td>7</td>
<td>0.08</td>
</tr>
<tr>
<td>0.40</td>
<td>5</td>
<td>0.08</td>
</tr>
<tr>
<td>0.50</td>
<td>4</td>
<td>0.06</td>
</tr>
</tbody>
</table>
• Trying to determine whether there is a dose-response relationship is a phase III objective.

• Using more than two dose levels to determine an OBD is even more ambitious.
Traditional Phase II Trials

• Estimate the proportion of tumors that shrink by 50% or more when the drug is administered either singly or in combination to patients with advanced stage tumors of a specific primary site
Objectives of Phase II Trials of Targeted Agents

• Determine whether there is a population of patients for whom the drug demonstrates sufficient anti-tumor activity to warrant a phase III trial
• Optimize the regimen in which the drug will be used in the phase III trial
• Optimize the target population for the phase III trial
• Develop tumor classifier for identifying target population and assay for reproducibly reading classifier
Endpoints for Phase II

• Tumor shrinkage
• Inhibition of target pathway
  – Ideally established in phase I
• Time to progression or proportion of patients without progression at a specified time
Optimal two-stage designs for phase II clinical trials

R Simon
Controlled Clinical Trials 10:1-10, 1989.
Optimal Two-Stage Phase II Designs

- Enter $n_1$ patients
- If response rate $\leq \frac{r_1}{n_1}$ reject drug
- Otherwise, enter $n_2$ additional patients
- If response rate $\leq \frac{r_2}{(n_1+n_2)}$ reject drug
Optimal Two-Stage Phase II Designs

• Given $p_0$, $p_1$, $\alpha$, and $\beta$
• Find $n_1$, $n_2$, $r_1$, $r_2$ to minimize
  – $E(\text{sample size} | p_0)$
  – $n_1 + n_2$
• $\Pr(\text{reject drug} | p_0) \geq 1 - \alpha$
• $\Pr(\text{reject drug} | p_1) \leq \beta$
• otsd.zip software at Statlib or linus.nci.nih.gov/brb
• On-line program at BRB website
Optimal Single Arm Two-Stage Design of Tumor Shrinkage

• To distinguish 5% \((p_0)\) response rate from 25% \((p_1)\) response rate with 10% false positive and false negative error rates:
  – Accrue 9 patients. Stop if no responses
  – If at least 1 response in first 9, continue accrual to 24 patients total
    • “Accept” treatment if at least 3/24 responses

• For regimens with 5% true response rate, the probability of stopping after 9 patients is 63%
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- Optimal Two-Stage Phase II Design Software
Sample Size Calculation for Randomized Clinical Trials

- Optimal Two-Stage Phase II Design

- Biomarker Targeted Randomized Design*
  1. Binary Outcome Endpoint
  2. Survival and Time-to-Event Endpoint

* Targeted design randomizes only marker positive patients to treatment or control arm. Untargeted design does not measure marker and randomizes all who otherwise are eligible.

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Patient Accrual in Phase II

• If the phase II trial for a particular primary site is not enriched for patients thought responsive to the drug, an initial stage of 10-15 patients may contain very few responsive patients.
  – Single stage design of 25-30 patients may be better

• Accrual of separate cohort of 25-30 patients whose tumors express target gives best chance to evaluate drug
Non-randomized Phase II Designs of Combinations

• Babe Ruth designs
• Difficult to interpret
  – Phase II designs with phase III objectives
• Thall-Simon Bayesian phase IIb designs using explicit controls to specify prior for control response rate


Using Time to Progression as Endpoint in Phase II Trials

• Requires comparison to distribution of progression times for patients not receiving drug

• Proportion of patients without progression at a specified time also requires comparison for evaluation

• Historical control vs randomized comparison
Time to Progression Endpoint

- It is difficult to reliably evaluate time to progression endpoint without a randomized control group.
- With historical controls, specific controls should be used for whom comparability of prognosis and surveillance for progression can be established.
Number of Patients on Experimental Treatment to have 80% Power for Detecting 15% Absolute Increase ($\alpha=.05$) in PFS vs Historical Controls

<table>
<thead>
<tr>
<th>Number of Historical Controls</th>
<th>90% Control Progression at landmark t</th>
<th>80% Control Progression at landmark t</th>
</tr>
</thead>
<tbody>
<tr>
<td>20</td>
<td>&gt;1000</td>
<td>&gt;1000</td>
</tr>
<tr>
<td>30</td>
<td>223</td>
<td>&gt;1000</td>
</tr>
<tr>
<td>40</td>
<td>108</td>
<td>285</td>
</tr>
<tr>
<td>50</td>
<td>80</td>
<td>167</td>
</tr>
<tr>
<td>75</td>
<td>58</td>
<td>101</td>
</tr>
<tr>
<td>100</td>
<td>50</td>
<td>83</td>
</tr>
<tr>
<td>200</td>
<td>42</td>
<td>65</td>
</tr>
</tbody>
</table>
Randomized Phase II Designs

- Randomized screening designs for selecting among new regimens
- Randomized discontinuation design
- Phase 2.5 design
- Phase 2/3 design
Randomized Phase II Screening Designs Using Biological Response

- For evaluating multiple new drugs or regimens to select most promising for further evaluation
  - Arm with greatest observed response rate is selected regardless of how small the difference is
  - Not for comparing a new drug/regimen to control

- Randomization better ensures uniform patient selection and evaluation

- Can be viewed as parallel optimum two-stage designs with randomization. Each arm evaluated as activity level $>p_1$ or $<p_0$
Patients per Arm for 2-arm Randomized Selection Design
Assures Correct Selection When True Response Probabilities Differ by 10%

<table>
<thead>
<tr>
<th>Response Probability of Inferior Rx</th>
<th>85% Probability of Correct Selection</th>
<th>90% Probability of Correct Selection</th>
</tr>
</thead>
<tbody>
<tr>
<td>5%</td>
<td>20</td>
<td>29</td>
</tr>
<tr>
<td>10%</td>
<td>28</td>
<td>42</td>
</tr>
<tr>
<td>20%</td>
<td>41</td>
<td>62</td>
</tr>
<tr>
<td>40%</td>
<td>54</td>
<td>82</td>
</tr>
</tbody>
</table>
Randomized Selection Design With Binary Endpoint

• K treatment arms
• n patients per arm
• Select arm with highest observed response rate
• $p_i = \text{true response probability for i’th arm}$
• $p_i = p_{\text{good}}$ with probability $\gamma$, otherwise $p_{\text{bad}}$
• With N total patients, determine K and n to maximize probability of finding a good rx
Probability of Selecting a Good Treatment
When $p_{\text{bad}}=0.1$, $p_{\text{good}}=0.5$ and $\gamma=0.1$

<table>
<thead>
<tr>
<th>$n$</th>
<th>$K$</th>
<th>Probability</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>20</td>
<td>0.626</td>
</tr>
<tr>
<td>10</td>
<td>10</td>
<td>0.590</td>
</tr>
<tr>
<td>15</td>
<td>7</td>
<td>0.511</td>
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<tr>
<td>20</td>
<td>5</td>
<td>0.414</td>
</tr>
<tr>
<td>25</td>
<td>4</td>
<td>0.344</td>
</tr>
</tbody>
</table>
Probability of Selecting a Good Treatment
When $p_{\text{bad}}=0.1$, $p_{\text{good}}=0.3$ and $\gamma=0.1$

<table>
<thead>
<tr>
<th>$n$</th>
<th>$K$</th>
<th>Probability</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>20</td>
<td>0.319</td>
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<td>10</td>
<td>0.375</td>
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<td>15</td>
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<td>0.383</td>
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<tr>
<td>20</td>
<td>5</td>
<td>0.341</td>
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<tr>
<td>25</td>
<td>4</td>
<td>0.309</td>
</tr>
</tbody>
</table>
Phase 2.5 Trial Design for Comparing New Regimen to Control Using PFS Endpoint


Phase 2.5 Trial Design

• Randomization to chemotherapy alone or with new drug
• Endpoint is progression free survival regardless of whether it is a validated surrogate of survival
• One-sided significance level can exceed .05 for analysis and sample size planning
Number of Events Required for Randomized Trial With Time to Event Endpoint

\[ E = 2 \left( \frac{k_\alpha + k_\beta}{\ln(\delta)} \right)^2 \]

\( \delta = \) hazard ratio or ratio of medians

For \( \alpha = 0.05, \beta = 0.20, \delta = 1.67 \) (40% reduction in hazard),
\( E = 47 \) events are required

For \( \alpha = 0.10 \), 35 events

For \( \alpha = 0.05, \beta = 0.20, \delta = 1.5 \), (33% reduction in hazard),
\( E = 75 \) events are required

For \( \alpha = 0.10 \), 55 events
Total Sample Size  
Randomized Phase 2.5  
2 years accrual, 1.5 years followup

<table>
<thead>
<tr>
<th>Improvement in median PFS</th>
<th>Hazard Ratio</th>
<th>$\alpha=0.05$</th>
<th>$\alpha=0.10$</th>
<th>$\alpha=0.20$</th>
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</thead>
<tbody>
<tr>
<td>4 → 6 months</td>
<td>1.5</td>
<td>216</td>
<td>168</td>
<td>116</td>
</tr>
<tr>
<td>6 → 9 months</td>
<td>1.5</td>
<td>228</td>
<td>176</td>
<td>120</td>
</tr>
<tr>
<td>4 → 8 months</td>
<td>2</td>
<td>76</td>
<td>60</td>
<td>40</td>
</tr>
<tr>
<td>6 → 12 months</td>
<td>2</td>
<td>84</td>
<td>64</td>
<td>44</td>
</tr>
</tbody>
</table>
Randomized Discontinuation Design (RDD) (Ratain et al.)

- The RDD starts all patients on the drug
- Patients with early progression go off study
- Patients with objective response continue on the drug
- Other patients are randomized to continue the drug or stop administration and be observed
- PFS from time of randomization is the endpoint
Randomized discontinuation design vs. upfront randomization

First stage -16 weeks, second stage - 16 weeks, progression is defined as a 120% increase from baseline. Overall sample size chosen to have 50 patients per arm in stage 2

<table>
<thead>
<tr>
<th>CAI Effect</th>
<th>Overall Sample size</th>
<th>Randomized discontinuation design</th>
<th>Upfront randomization</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>First stage SD rate at 16 weeks</td>
<td>Second stage PD rate at 32 weeks CAI/placebo</td>
</tr>
<tr>
<td>0</td>
<td>333</td>
<td>.30</td>
<td>.73/.73</td>
</tr>
<tr>
<td>.1</td>
<td>290</td>
<td>.345</td>
<td>.70/.735</td>
</tr>
<tr>
<td>.2</td>
<td>250</td>
<td>.40</td>
<td>.67/.74</td>
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<tr>
<td>.3</td>
<td>216</td>
<td>.47</td>
<td>.63/.75</td>
</tr>
<tr>
<td>.4</td>
<td>184</td>
<td>.54</td>
<td>.59/.76</td>
</tr>
<tr>
<td>.5</td>
<td>160</td>
<td>.63</td>
<td>.53/.76</td>
</tr>
<tr>
<td>.6</td>
<td>138</td>
<td>.73</td>
<td>.45/.77</td>
</tr>
</tbody>
</table>
Comparison of Designs Under Modified Model of Treatment Effect

Treatment is assumed to have no effect for patients with rapidly growing tumors; i.e., tumors which would grow untreated by more than cutoff % at 16 weeks.

<table>
<thead>
<tr>
<th>CAI Effect</th>
<th>Cutoff</th>
<th>Overall Sample size</th>
<th>Randomized discontinuation design</th>
<th>Upfront randomization</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>First stage SD rate at 16 weeks</td>
<td>Second stage PD rate at 32 weeks CAI/placebo</td>
</tr>
<tr>
<td>.4</td>
<td>20%</td>
<td>184</td>
<td>.54</td>
<td>.59/.76</td>
</tr>
<tr>
<td>.4</td>
<td>17%</td>
<td>184</td>
<td>.54</td>
<td>.59/.76</td>
</tr>
<tr>
<td>.4</td>
<td>13%</td>
<td>184</td>
<td>.54</td>
<td>.59/.76</td>
</tr>
<tr>
<td>.6</td>
<td>30%</td>
<td>138</td>
<td>.73</td>
<td>.45/.77</td>
</tr>
<tr>
<td>.6</td>
<td>23%</td>
<td>138</td>
<td>.73</td>
<td>.45/.77</td>
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<tr>
<td>.6</td>
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<td>138</td>
<td>.73</td>
<td>.45/.77</td>
</tr>
<tr>
<td>.6</td>
<td>15%</td>
<td>138</td>
<td>.73</td>
<td>.45/.77</td>
</tr>
</tbody>
</table>
Randomized Discontinuation Design (RDD)

- The RDD can facilitate observing an effect of the drug on PFS compared to a standard randomized phase 2.5 design
  - The RDD requires a large sample size
  - The RDD is not a phase III trial because it does not establish the clinical utility of administering the drug to the patient compared to not administering it
Phase II/III Design

• Randomized trial comparing regimen containing new drug to control regimen
• Perform interim analysis comparing treatments using PFS (progression-free survival) endpoint
• If $p_{pfs} < p^*$ then continue trial to evaluate phase III endpoint
• Otherwise, terminate trial
• Conducting a phase III trial in the traditional way with tumors of a specified site/stage/pre-treatment category may result in a false negative trial
  – Unless a sufficiently large proportion of the patients have tumors driven by the targeted pathway
• Positive results in phase III trials may be driven by a subset of patients whose tumors are driven by the targeted pathway
  – Such trials may result in treatment of the majority with very expensive drugs for the benefit of the minority
• It is important to characterize in phase II studies which tumors are most likely to be sensitive to the drug
Strategies for Development of Genomic Classifiers

• Type of Classifier
  – Single gene or protein based on knowledge of therapeutic target
    • HER2 amplification
    • EGFR mutation or amplification
    – Empirically determined based on correlating gene expression or genotype to patient outcome after treatment.

• When to develop Classifier
  – During phase II
  – After failed phase III trial or after broad indication drug approval
Single gene or protein based on knowledge of therapeutic target

- Often there will be several assays that can be used
- Phase II development is a good time to establish which assay should be used in phase III
- The assay to be used for either selecting patients for phase III trial or for testing hypotheses in the phase III trial should be determined before starting the phase III trial
• Refining the target based single gene or protein assay to use in phase III can be accomplished either from traditional single arm phase II trials with response endpoint or from randomized phase 2.5 trials with PFS endpoint
Guiding Principle

• The data used to develop the classifier must be distinct from the data used to test hypotheses about treatment effect in subsets determined by the classifier
  – Developmental studies are exploratory
  – Studies on which treatment effectiveness claims are to be based should be definitive hypothesis testing studies based on completely pre-specified classifiers
Development of Empirical Gene Expression Based Classifier

• 20 phase II responders are needed to compare to non-responders in order to develop signature for predicting response
  – Dobbin KK, Simon RM. Sample size planning for developing classifiers using high dimensional DNA microarray data, Biostatistics (In Press); available at http://linus.nci.nih.gov
Summary

• Phase I trials of molecularly targeted agents should generally be based on determining a dose that can be delivered repeatedly over time with acceptable toxicity and then evaluating whether the targeted pathway is inhibited at that dose.

• Phase II trials are most efficient if anti-tumor effect can be evaluated in individual tumors rather than use of PFS.
Summary

• Phase II trials should include establishment and refinement of classifiers of which tumors are most likely to respond
• Such classifiers should be completely specified prior to launching of phase III trials
• Phase III trials should incorporate prospectively specified plans for the utilization of biomarker classifiers of patients with sensitive tumors
Moving from Correlative Studies to Predictive Medicine

Richard Simon, D.Sc.
Chief, Biometric Research Branch
National Cancer Institute
http://linus.nci.nih.gov/brb
Biomarker

• “Any biological measurement that provides actionable information regarding disease progression, pharmacology, or safety that can be used as a basis for decision making in drug development.”
  – J. Boguslavsky
• “I don’t know what ‘clinical validation’ [of a biomarker] means. The first thing you have to do is define a purpose for the biomarker. Validation is all about demonstrating fitness for purpose.”

– Dr. Stephen Williams, Pfizer
Biomarkers

• Surrogate endpoints
  – A measurement made on a patient before, during and after treatment to determine whether the treatment is working

• Predictive classifier
  – A measurement made before treatment to predict whether a particular treatment is likely to be beneficial
Surrogate Endpoints

• It is extremely difficult to properly validate a biomarker as a surrogate for clinical outcome. It requires a series of randomized trials with both the candidate biomarker and clinical outcome measured.
Cardiac Arrhythmia Suppression Trial

- Ventricular premature beats was proposed as a surrogate for survival
- Antiarrythmic drugs supressed ventricular premature beats but killed patients at approximately 2.5 times that of placebo
• It is rare that we understand disease pathophysiology well enough to argue that a biomarker is self evidently a proper surrogate endpoint for clinical utility
• It is often more difficult and time consuming to properly “validate” an endpoint as a surrogate than to use the clinical endpoint in phase III trials
• The time frame for validating a surrogate is inconsistent with the time frame for initiating a pivotal study
Surrogate Endpoints

• It is often more difficult to properly “validate” a surrogate than to use the clinical endpoint in phase III trials
Using Intermediate Endpoints Not Established as Surrogates of Clinical Benefit

• Biomarkers can be useful in phase I/II studies and need not be validated as surrogates for clinical outcome

• Unvalidated surrogates can also be used for early termination of phase III trials. The trial should continue accrual and follow-up to evaluate true endpoint if treatment effect on partial surrogate is sufficient
Biomarkers for Treatment Selection

- Oncologists need improved tools for selecting treatment for individual patients.
- Most cancer treatments benefit only a minority of patients to whom they are administered.
- Being able to predict which patients are likely to benefit would save patients from unnecessary toxicity, inconvenience and enhance their chance of receiving a drug that helps them.
- The current over treatment of patients results in a major expense for individuals and society.
Oncology Needs Predictive Markers not Prognostic Factors

• Most prognostic factors are not used because they are not therapeutically relevant

• Most prognostic factor studies use a convenience sample of patients for whom tissue is available. Generally the patients are too heterogeneous to support therapeutically relevant conclusions
Pusztai et al. The Oncologist 8:252-8, 2003

- 939 articles on “prognostic markers” or “prognostic factors” in breast cancer in past 20 years
- ASCO guidelines only recommend routine testing for ER, PR and HER-2 in breast cancer
- “With the exception of ER or progesterone receptor expression and HER-2 gene amplification, there are no clinically useful molecular predictors of response to any form of anticancer therapy.”
Predictive Classifiers

• Most cancer treatments benefit only a minority of patients to whom they are administered
  – Particularly true for molecularly targeted drugs
• Being able to predict which patients are likely to benefit would
  – save patients from unnecessary toxicity, and enhance their chance of receiving a drug that helps them
  – Help control medical costs
“If new refrigerators hurt 7% of customers and failed to work for another one-third of them, customers would expect refunds.”

• Clinical trial for patients with breast cancer, without nodal or distant metastases, Estrogen receptor positive tumor
  – 5 year survival rate for control group (surgery + radiation + Tamoxifen) expected to be 90%
  – Size trial to detect 92% survival in group treated with control modalities plus chemotherapy
Treating the Many for the Benefit of the Few

- Acceptable to industry
- Acceptable to statisticians
  - Broad eligibility
    - Uncertainty principle
  - Avoid subset analysis
    - “Do it but don’t believe it”
- Convenient for treating physician
- Not so good for patients or for their health budget
Disease-Free Survival

P = 0.002

- Tam 771
- MFT 767
- CMFT 768

Events

Year

0 1 2 3 4 5

% 90 80 70

- Al 753 723 688 628 461
- Risk 757

* Comparison to Tamoxifen

Relative Risk (95% Confidence Interval)

- MFT/Tam 0.72 (0.56-0.93)
- CMFT/Tam 0.65 (0.50-0.84)

Distant Disease-Free Survival

P = 0.005

- Events 115
- P* 0.003

Year

0 1 2 3 4 5

% 90 80 70

- Al 756 731 703 642 491
- Risk 757

- MFT 737 713 658 505
- CMFT 763 93 0.001

Survival

P = 0.04

- Deaths 60
- P* 0.05

Year

0 1 2 3 4 5

% 90 80 70

- Al 771 767 750 592 531
- Risk 756 740 719 672 493

- MFT 766 760 742 701 542
- CMFT 764 530

RELATIVE RISK (95% CONFIDENCE INTERVAL)

- MFT/Tam 0.68 (0.51-0.90)
- CMFT/Tam 0.67 (0.50-0.89)
- MFT/Tam 0.64 (0.42-0.95)
“Hypertension is not one single entity, neither is schizophrenia. It is likely that we will find 10 if we are lucky, or 50, if we are not very lucky, different disorders masquerading under the umbrella of hypertension. I don’t see how once we have that knowledge, we are not going to use it to genotype individuals and try to tailor therapies, because if they are that different, then they’re likely fundamentally … different problems…”

– George Poste
Conventional Broad Eligibility Phase III Trials May Result In

- Large trials with false negative results because the proportion of patients who benefit is too small to provide adequate statistical power
- Statistically significant outcome that results in subsequent treatment of many patients who don’t benefit
• Targeted clinical trials can be much more efficient than untargeted clinical trials, if we know who to target
• In new drug development, the role of a classifier is to select a target population for treatment
  – The focus should be on evaluating the new drug in a population defined by a predictive classifier, not on “validating” the classifier
• FDA criteria for validation of surrogate endpoints should not be applied to predictive classifiers
There Should Be No Requirement For

• Demonstrating that the classifier or any of its components are “validated biomarkers of disease status”

• Ensuring that the individual components of the classifier are correlated with patient outcome or effective for selecting patients for treatment

• Demonstrating that repeating the classifier development process on independent data results in the same classifier
One Should Require That

- The classifier be reproducibly measurable
- The classifier in conjunction with the medical product has clinical utility
Biomarker validation vs pharmacogenomic classifier utilization

• Adoption of a pharmacogenomic classifier to restrict the use of a treatment in wide use should be based on adequate validation of the classifier
  – Validation means demonstrating that the classifier leads to better clinical outcome
• In new drug development, the role of a classifier is to select a target population for treatment
  – The focus should be on evaluating the new drug, not on validating the classifier
Developmental Strategy (I)

- **Develop** a diagnostic classifier that identifies the patients likely to benefit from the new drug
- Develop a reproducible assay for the classifier
- **Use** the diagnostic to restrict eligibility to a prospectively planned evaluation of the new drug
- Demonstrate that the new drug is effective in the prospectively defined set of patients determined by the diagnostic
Using phase II data, develop predictor of response to new drug.

Develop Predictor of Response to New Drug

Patient Predicted Responsive
  - New Drug
  - Control

Patient Predicted Non-Responsive
  - Off Study
Evaluating the Efficiency of Strategy (I)


• reprints and interactive sample size calculations at http://linus.nci.nih.gov/brb
Pharmacogenomic Model for Two Treatments With Binary Response

• Molecurally targeted treatment E
• Control treatment C
• $\gamma$ Proportion of patients that express target
• $p_c$ control response probability
• response probability for E patients who express target is $(p_c + \delta_1)$
• Response probability for E patients who do not express target is $(p_c + \delta_0)$
Approximations

• Observed response rate $\sim N(p,p(1-p)/n)$

• $p_e(1-p_e) \sim p_c(1-p_c)$
Two Clinical Trial Designs

• Un-targeted design
  – Randomized comparison of E to C without screening for expression of molecular target

• Targeted design
  – Assay patients for expression of target
  – Randomize only patients expressing target
Number of Randomized Patients Required

• Type I error $\alpha$
• Power $1-\beta$ for obtaining significance

\[ n = 2(p_c q_c + p_e q_e) \left( \frac{k_{1-\alpha} + k_{1-\beta}}{p_e - p_c} \right)^2 \]
• For targeted design
  \(- p_e = p_c + \delta_1 \)
  \(- p_e - p_c = \delta_1 \)

• For un-targeted design
  \(- p_e = (1 - \gamma)(p_c + \delta_0) + \gamma(p_c + \delta_1) \)
  \(- p_e - p_c = \gamma \delta_1 + (1 - \gamma) \delta_1 \)
Randomized Ratio
(normal approximation)

- RandRat = \( \frac{n_{\text{untargeted}}}{n_{\text{targeted}}} \)

\[
RandRat \approx \left( \frac{\delta_1}{\gamma \delta_1 + (1 - \gamma) \delta_0} \right)^2
\]

- \( \delta_1 = \) rx effect in marker + patients
- \( \delta_0 = \) rx effect in marker - patients
- \( \gamma = \) proportion of marker + patients

- If \( \delta_0 = 0 \), RandRat = \( 1/\gamma^2 \)
- If \( \delta_0 = \delta_1/2 \), RandRat = \( 4/(\gamma + 1)^2 \)
### Randomized Ratio

\[ \frac{n_{\text{untargeted}}}{n_{\text{targeted}}} \]

<table>
<thead>
<tr>
<th>Proportion Assay Positive</th>
<th>No Treatment Benefit for Assay Negative Patients</th>
<th>Treatment Benefit for Assay Negative Patients is Half That for Assay Positive Patients</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.75</td>
<td>1.78</td>
<td>1.31</td>
</tr>
<tr>
<td>0.5</td>
<td>4</td>
<td>1.78</td>
</tr>
<tr>
<td>0.25</td>
<td>16</td>
<td>2.56</td>
</tr>
</tbody>
</table>
## Screened Ratio

<table>
<thead>
<tr>
<th>$\gamma$</th>
<th>Assay +</th>
<th>$\delta_0 = 0$</th>
<th>$\delta_0 = \delta_1 / 2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.75</td>
<td></td>
<td>1.33</td>
<td>0.98</td>
</tr>
<tr>
<td>0.5</td>
<td></td>
<td>2</td>
<td>0.89</td>
</tr>
<tr>
<td>0.25</td>
<td></td>
<td>4</td>
<td>0.64</td>
</tr>
</tbody>
</table>
Imperfect Assay Sensitivity & Specificity

• $\lambda_{\text{sens}}$ = sensitivity
  – $\Pr[\text{assay}+ | \text{target expressed}]$

• $\lambda_{\text{spec}}$ = specificity
  – $\Pr[\text{assay}- | \text{target not expressed}]$
Proportion of Assay Positive Patients That Express Target

\[ w_1 = \frac{\gamma \lambda_{sens}}{\gamma \lambda_{sens} + (1 - \gamma)(1 - \lambda_{spec})} \]

<table>
<thead>
<tr>
<th>( \lambda_{sens} )</th>
<th>( \lambda_{spec} )</th>
<th>( \gamma )</th>
<th>( w_1 )</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.9</td>
<td>0.9</td>
<td>0.75</td>
<td>0.96</td>
</tr>
<tr>
<td>0.9</td>
<td>0.9</td>
<td>0.5</td>
<td>0.9</td>
</tr>
<tr>
<td>0.9</td>
<td>0.9</td>
<td>0.25</td>
<td>0.75</td>
</tr>
<tr>
<td>0.9</td>
<td>0.9</td>
<td>0.10</td>
<td>0.50</td>
</tr>
</tbody>
</table>
Randomized Ratio

- $\text{RandRat} = \frac{n_{\text{untargeted}}}{n_{\text{targeted}}}$

$$\text{RandRat} = \left( \frac{w_1 \delta_1 + (1 - w_1) \delta_0}{\gamma \delta_1 + (1 - \gamma) \delta_0} \right)^2$$
Randomized Ratio
sensitivity=specificity=0.9

<table>
<thead>
<tr>
<th>$\gamma$</th>
<th>$\delta_0=0$</th>
<th>$\delta_0=\delta_1/2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Express target</td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.75</td>
<td>1.29</td>
<td>1.26</td>
</tr>
<tr>
<td>0.5</td>
<td>1.8</td>
<td>1.6</td>
</tr>
<tr>
<td>0.25</td>
<td>3.0</td>
<td>1.96</td>
</tr>
<tr>
<td>0.1</td>
<td>25.0</td>
<td>1.86</td>
</tr>
</tbody>
</table>
Screened Ratio

Imperfect Assay

- \( N_{\text{untargeted}} = n_{\text{untargeted}} \)

\[ N_{\text{targeted}} = \frac{n_{\text{targeted}}}{\gamma \lambda_{\text{sens}} + (1 - \gamma)(1 - \lambda_{\text{spec}})} \]

\[ \text{ScreenRat} = \left[ \gamma \lambda_{\text{sens}} + (1 - \gamma)(1 - \lambda_{\text{spec}}) \right] R_{\text{andra}} \]
Screened Ratio
sensitivity=specificity=0.9

<table>
<thead>
<tr>
<th>$\gamma$</th>
<th>$\delta_0 = 0$</th>
<th>$\delta_0 = \delta_1 / 2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Express target</td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.75</td>
<td>0.9</td>
<td>0.88</td>
</tr>
<tr>
<td>0.5</td>
<td>0.9</td>
<td>0.80</td>
</tr>
<tr>
<td>0.25</td>
<td>0.9</td>
<td>0.59</td>
</tr>
<tr>
<td>0.1</td>
<td>4.5</td>
<td>0.33</td>
</tr>
</tbody>
</table>
• For Trastuzumab, even a relatively poor assay enabled conduct of a targeted phase III trial which was crucial for establishing effectiveness

• Recent results with Trastuzumab in early stage breast cancer show dramatic benefits for patients selected to express Her-2
## Comparison of Targeted to Untargeted Design

Simon R, Development and Validation of Biomarker Classifiers for Treatment Selection, JSPI

<table>
<thead>
<tr>
<th>Treatment Hazard Ratio for Marker Positive Patients</th>
<th>Number of Events for Targeted Design</th>
<th>Number of Events for Traditional Design</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Percent of Patients Marker Positive</td>
</tr>
<tr>
<td></td>
<td></td>
<td>20% 33% 50%</td>
</tr>
<tr>
<td>0.5</td>
<td>74</td>
<td>2040 720 316</td>
</tr>
</tbody>
</table>
Interactive Software for Evaluating a Targeted Design

• http://linus.nci.nih.gov/brb/
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BRB Alumni

Sample Size Calculation

BRB Annual Report 2005

Mathematics and Oncology
- The Norton-Simon Hypothesis
- The Norton-Simon Hypothesis and Breast Cancer Mortality in National Randomized Trial

Position Available
Post-doctoral fellow positions available

Software Download
- Accelerated Titration Design Software
- Optimal Two-Stage Phase II Design Software
Sample Size Calculation for Randomized Clinical Trials

- Optimal Two-Stage Phase II Design

- Biomarker Targeted Randomized Design*
  1. Binary Outcome Endpoint
  2. Survival and Time-to-Event Endpoint

* Targeted design randomizes only marker positive patients to treatment or control arm. Untargeted design does not measure marker and randomizes all who otherwise are eligible.

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Sample Size Calculation: Binary Outcome Endpoint


\[
\begin{align*}
pc & \\
gamma & \\
delta_1 & \\
delta_0 & \\
alpha & 0.05 \\
power & 0.90
\end{align*}
\]

Submit

\(pc\) = probability of "response" for control arm

\(gamma\) = proportion of patients who are classifier negative (i.e. less responsive to new treatment)

\(delta_1\) = improvement in response probability for new treatment in classifier positive patients

\(delta_0\) = improvement in response probability for new treatment in classifier negative patients

\(alpha\) = two-sided significance level

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Developmental Strategy (II)

Develop Predictor of Response to New Rx

Predicted Responsive To New Rx
- New RX
- Control

Predicted Non-responsive to New Rx
- New RX
- Control
Developmental Strategy (II)

• Do not use the diagnostic to restrict eligibility, but to structure a prospective analysis plan.

• Compare the new drug to the control overall for all patients ignoring the classifier.
  – If $p_{\text{overall}} \leq 0.04$ claim effectiveness for the eligible population as a whole

• Otherwise perform a single subset analysis evaluating the new drug in the classifier + patients
  – If $p_{\text{subset}} \leq 0.01$ claim effectiveness for the classifier + patients.
Key Features of Design (II)

• The purpose of the RCT is to evaluate treatment T vs C overall and for the pre-defined subset; not to re-evaluate the components of the classifier, or to modify or refine the classifier
Sample Size Planning for Design II

1. Size for standard power (e.g. 0.9) for detecting usual treatment effect $d$ (e.g. 15%) at significance level 0.04

2. Size for standard power (e.g. 0.9) for detecting treatment effect in subset of size $d / \text{proportion positive}$

3. Size as in 1 but extend accrual of classifier positive patients if overall test is non-significant
Developmental Strategy (IIb)

- Do not use the diagnostic to restrict eligibility, but to structure a prospective analysis plan.
- Compare the new drug to the control for classifier positive patients
  - If $p_+ > 0.05$ make no claim of effectiveness
  - If $p_+ \leq 0.05$ claim effectiveness for the classifier positive patients and
    - Continue accrual of classifier negative patients and eventually test treatment effect at 0.05 level
Sample size Planning for IIb

- Accrue classifier positive and negative patients until there are sufficient classifier positive patients for standard power at significance level 0.05 for detecting large treatment effect $D$
- If treatment is found effective in classifier + patients, continue accrual of negative patients for standard power at significance level 0.05 for detecting usual size treatment effect $d$ representing minimal useful clinical utility
The Roadmap

1. Develop a completely specified genomic classifier of the patients likely to benefit from a new drug
2. Establish reproducibility of measurement of the classifier
3. Use the completely specified classifier to design and analyze a new clinical trial to evaluate effectiveness of the new treatment with a pre-defined analysis plan.
Guiding Principle

• The data used to develop the classifier must be distinct from the data used to test hypotheses about treatment effect in subsets determined by the classifier
  – Developmental studies are exploratory
  – Studies on which treatment effectiveness claims are to be based should be definitive studies that test a treatment hypothesis in a patient population completely pre-specified by the classifier
Use of Archived Samples

• From a non-targeted “negative” clinical trial to develop a binary classifier of a subset thought to benefit from treatment

• Test that subset hypothesis in a separate clinical trial
  – Prospective targeted type (I) trial
  – Prospective type (II) trial
  – Using archived specimens from a second previously conducted clinical trial
Development of Genomic Classifiers

• Single gene or protein based on knowledge of therapeutic target
• Single gene or protein culled from set of candidate genes identified based on imperfect knowledge of therapeutic target
• Empirically determined based on correlating gene expression to patient outcome after treatment
Development of Genomic Classifiers

• During phase II development or

• After failed phase III trial using archived specimens.

• Adaptively during early portion of phase III trial.
Development of Empirical Gene Expression Based Classifier

• 20-30 phase II responders are needed to compare to non-responders in order to develop signature for predicting response
  – Dobbin KK, Simon RM. Sample size planning for developing classifiers using high dimensional DNA microarray data, Biostatistics (In Press); available at http://linus.nci.nih.gov
Development of Empirical Gene Expression Based Classifier

• A signature of response to the new drug may not represent a signature of preferential benefit from a regimen containing the new drug versus a control regimen
Adaptive Signature Design
An adaptive design for generating and prospectively testing a gene expression signature for sensitive patients

Boris Freidlin and Richard Simon
Clinical Cancer Research 11:7872-8, 2005
Adaptive Signature Design
End of Trial Analysis

• Compare E to C for all patients at significance level 0.04
  – If overall H₀ is rejected, then claim effectiveness of E for eligible patients
  – Otherwise
• Otherwise:
  – Using only the first half of patients accrued during the trial, develop a binary classifier that predicts the subset of patients most likely to benefit from the new treatment E compared to control C
  – Compare E to C for patients accrued in second stage who are predicted responsive to E based on classifier
    • Perform test at significance level 0.01
    • If $H_0$ is rejected, claim effectiveness of E for subset defined by classifier
Treatment effect restricted to subset. 10% of patients sensitive. Sensitivity genes are uncorrelated. 400 patients, 10,000 genes.
Treatment effect restricted to subset. 10% of patients sensitive.

Sensitivity genes are correlated, 400 patients, 10,000 genes.
Treatment effect restricted to subset.
10% of patients sensitive, 10 sensitivity genes, 10,000 genes, 400 patients.

<table>
<thead>
<tr>
<th>Test</th>
<th>Power</th>
</tr>
</thead>
<tbody>
<tr>
<td>Overall .05 level test</td>
<td>46.7</td>
</tr>
<tr>
<td>Overall .04 level test</td>
<td>43.1</td>
</tr>
<tr>
<td>Sensitive subset .01 level test</td>
<td>42.2</td>
</tr>
<tr>
<td>(performed only when overall .04 level test is negative)</td>
<td></td>
</tr>
<tr>
<td>Overall adaptive signature design</td>
<td>85.3</td>
</tr>
</tbody>
</table>
Overall treatment effect, no subset effect.
10,000 genes, 400 patients.

<table>
<thead>
<tr>
<th>Test</th>
<th>Power</th>
</tr>
</thead>
<tbody>
<tr>
<td>Overall .05 level test</td>
<td>74.2</td>
</tr>
<tr>
<td>Overall .04 level test</td>
<td>70.9</td>
</tr>
<tr>
<td>Sensitive subset .01 level test</td>
<td>1.0</td>
</tr>
<tr>
<td>Overall adaptive signature design</td>
<td>70.9</td>
</tr>
</tbody>
</table>
Validation of Predictive Classifiers for Use with Available Treatments

• Should establish that the classifier is robust, reproducibly measurable and has clinical utility

• Studies of predictive classifiers should be viewed as either developmental or validation studies
Studies Developing Gene Expression Profile Classifiers Should be Viewed as Analogous to Phase II Trials Requiring Phase III Validation
Developmental Studies

• Develop classifier that either predicts outcome of patients receiving specified treatment or control treatment

• Uses split-sample validation or cross-validation to estimate predictive accuracy of classifier
Split-Sample Evaluation

• Training-set
  – Used to select features, select model type, determine parameters and cut-off thresholds

• Test-set
  – Withheld until a single model is fully specified using the training-set.
  – Fully specified model is applied to the expression profiles in the test-set to predict class labels.
  – Number of errors is counted
  – Ideally test set data is from different centers than the training data and assayed at a different time
Non-Cross-Validated Prediction

1. Prediction rule is built using full data set.
2. Rule is applied to each specimen for class prediction.

Cross-Validated Prediction (Leave-One-Out Method)

1. Full data set is divided into training and test sets (test set contains 1 specimen).
2. Prediction rule is built from scratch using the training set.
3. Rule is applied to the specimen in the test set for class prediction.
4. Process is repeated until each specimen has appeared once in the test set.
• Cross validation is only valid if the test set is not used in any way in the development of the model. Using the complete set of samples to select genes violates this assumption and invalidates cross-validation.

• With proper cross-validation, the model must be developed from scratch for each leave-one-out training set. This means that feature selection must be repeated for each leave-one-out training set.
  

• The cross-validated estimate of misclassification error is an estimate of the prediction error for model fit using specified algorithm to full dataset
Myth

• Split sample validation is superior to LOOCV or 10-fold CV for estimating prediction error
PREDICTION ERROR ESTIMATION: A COMPARISON OF RESAMPLING METHODS

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ABSTRACT

Motivation: In genomic studies, thousands of features are collected on relatively few samples. One of the goals of these studies is to build classifiers to predict the outcome of future observations. There are three inherent steps to this process: feature selection, model selection, and prediction assessment. With a focus on prediction assessment, we compare several methods for estimating the "true" prediction error of a prediction model in the presence of feature selection.

Results: For small studies where features are selected from thousands of candidates, the resubstitution and simple split-sample estimates are seriously biased. In these small samples, leave-one-out (LOOCV), 10-fold cross-validation (CV), and the .632+ bootstrap have the smallest bias for univariate discriminant analysis, nearest neighbor, and classification trees. LOOCV and 10-fold CV have the smallest bias for linear discriminant analysis. Additionally, LOOCV, 5- and 10-fold CV, and the .632+ bootstrap have the lowest mean square error. The .632+ bootstrap is quite biased in small sample sizes with strong signals to noise ratios. Differences in performance among resampling methods are reduced as the number of specimens available increases.

Availability: A complete compilation of results in tables and figures is available in Molinaro et al. (2005). A code for simulations and analyses is available from the authors.

Contact: annette.molinaro@yale.edu

1 INTRODUCTION

In genomic studies one frequently encounters high dimensional data and small sample sizes. Microarrays simultaneously monitor expression levels for several thousands of genes. Proteomic profiling studies using SELDI-TOF (surface-enhanced laser desorption and ionization time-of-flight) measure size and charge of proteins and protein fragments by mass spectrometry, and result in up to 15,000 intensity levels at pre-specified mass values for each specimen. Sample sizes in such experiments are typically less than 109.

In many studies observations are known to belong to predetermined classes and the task is to build predictors or classifiers for new observations whose class is unknown. Deciding which genes or protein measurements to include in the prediction is called feature selection and is a crucial step in developing a class predictor. Including too many noisy variables reduces accuracy of the prediction and may lead to over-fitting of data, resulting in promising but often non-reproducible results (Ramaswamy, 2004).

Another difficulty is model selection with numerous classification models available. An important step in reporting results is assessing the chosen model's error rate, or generalizability. In the absence of independent validation data, a common approach to estimating predictive accuracy is based on some form of resampling the original data, e.g., cross-validation. These techniques divide the data into a learning set and a test set and range in complexity from the popular learning-test split to 5-fold cross-validation, Monte-Carlo 5-fold cross-validation, and bootstrap resampling. Few comparisons of standard resampling methods have been performed so far, and all of them exhibit limitations that make their conclusions inapplicable to most genomic settings. Early comparisons of resampling techniques in the literature are focused on model selection as opposed to prediction error estimation (Breslow and Storer, 1992; Hynan, 1989). In two recent assessments of resampling techniques for error estimation (Stingo-Neto and Dougherty, 2004; Elton, 2004), feature selection was not included as part of the resampling procedures, causing the conclusions to be inappropriate for the high-dimensional setting.

We have performed an extensive comparison of resampling methods to estimate prediction error using simulated (large signal to noise ratio), microarray (intermediate signal to noise ratio) and proteomic data (low signal to noise ratio), encompassing increasing sample sizes with large numbers of features. The impact of feature selection on the performance of various error validation methods is highlighted. The results elucidate the 'best' resampling techniques for
• Both split-sample validation and cross-validation represent *internal validation*
Limitations to Internal Validation

- Sample handling and assay conduct are performed under controlled conditions that do not incorporate real world sources of variability
- Developmental studies are generally small
- Predictive accuracy is generally not clinical utility
External Validation

• From different clinical centers
• Specimens assayed at different time from training data
• Reproducibility of assay for individual tumors demonstrated to clinical reference laboratory standards
• Positive and negative samples collected in the same way
• Study addresses clinical utility of using the genomic classifier compared to using standard practice guidelines
Myth

• Development of good predictive classifiers is not possible with >1000 genes and <100 cases or requires huge sample sizes

• Predictive models should be reproducible on independent data
• Much of the conventional wisdom of statistical analysis is focused on inference, not on prediction
• Demonstrating statistical significance of prognostic factors is not the same as demonstrating predictive accuracy
• Predictive models should predict accurately for independent data; the model itself need not be reproducibly derivable on independent data
• Most statistical methods were not developed for prediction problems and particularly not for prediction problems with >10,000 variables and <100 cases
Concordance among Gene-Expression-Based Predictors for Breast Cancer

Cheng Fan, M.S., Daniel S. Oh, Ph.D., Lodewyk Wessels, Ph.D., Britta Wegelt, Ph.D., Dimitry S.A. Nuyten, M.D., Andrew B. Nobel, Ph.D., Laura van 't Veer, Ph.D., and Charles M. Perou, Ph.D.

ABSTRACT

BACKGROUND
Gene-expression-profiling studies of primary breast tumors performed by different laboratories have resulted in the identification of a number of distinct prognostic profiles, or gene sets, with little overlap in terms of gene identity.

METHODS
To compare the predictions derived from these gene sets for individual samples, we obtained a single data set of 295 samples and applied five gene-expression-based models: intrinsic subtypes, 70-gene profile, wound response, recurrence score, and the two-gene ratio for patients who had been treated with tamoxifen.

RESULTS
We found that most models had high rates of concordance in their outcome predictions for the individual samples. In particular, almost all tumors identified as having an intrinsic subtype of basal-like, HER2-positive and estrogen-receptor-negative, or luminal B (associated with a poor prognosis) were also classified as having a poor 70-gene profile, activated wound response, and high recurrence score. The 70 gene and recurrence-score models, which are beginning to be used in the clinical setting, showed 77 to 81 percent agreement in outcome classification.

CONCLUSIONS
Even though different gene sets were used for prognostication in patients with breast cancer, four of the five tested showed significant agreement in the outcome predictions for individual patients and are probably tracking a common set of biologic phenotypes.
Sample Size Planning

References

• K Dobbin, R Simon. Sample size determination in microarray experiments for class comparison and prognostic classification. Biostatistics 6:27-38, 2005

• K Dobbin, R Simon. Sample size planning for developing classifiers using high dimensional DNA microarray data. Biostatistics (In Press)
Clinical Targeting of Treatment to Cancer Patient Based on Tumor Expression Profile in Broad Clinical Use
Limited by Appropriate Therapeutic Decision Contexts

- Patients whose prognosis is so good without chemotherapy that it can be withheld
- Multiple effective treatments exist and need guidance in choosing among them
- Multiple palliative treatments exist
Limited by Appropriate Therapeutic Decision Contexts

• Patients whose prognosis is so good without chemotherapy that it can be withheld
  – Unwillingness of physicians to withhold treatment even if it’s chance for benefiting the patient is low
Limited by Appropriate Therapeutic Decision Contexts

• Potentially curative treatment for life threatening disease with no good alternative therapy
  – Not many curative treatments
  – Can rarely be sure that NPV is perfect
Developing Predictive Classifiers for Use with Existing Treatments

• Lack of financial incentives
• Difficulty in performing prospective validation studies that establish clinical utility
• Difficulty in establishing assay robustness and need for research-commercial partnership
• Limitations in practicality of existing platforms
Genomic Approach to Diagnostic/Prognostic Marker Development

• Select therapeutically relevant population
  – Node negative, ER+, well staged breast cancer patients who have received Tam alone and have long follow-up
• Perform genome wide expression profiling
• Develop multi-gene/protein predictor of outcome
• Obtain internal estimate of prediction accuracy
• Adapt platform to clinical application
• Establish assay reproducibility
• Conduct prospective study to establish clinical utility
Validation Study
Node negative Breast Cancer

- Prospective study design
- Samples collected and assayed from patients with node negative ER+ breast cancer who will receive TAM
- Apply single, fully specified multi-gene predictor of outcome to samples and categorize each patient as good or poor prognosis
- Categorizing each patient with regard to practice standards as requiring or not requiring chemotherapy
- Randomizing patients predicted to be poor prognosis by classifier for whom practice standards do not recommend chemotherapy
- Are long-term outcomes for randomized patients
Determine marker based rx (M-rx) and standard of care based rx (SOC-rx)

\[ M-rx = SOC-rx? \]

- Yes
  - Off study

- No
  - Randomize
    - M-rx
    - SOC-rx
Approximate number of events required for 80% power with 5% two-sided log-rank test for comparing arms of design shown in Figure 3. Only marker + patients are randomized. Treatment hazard ratio for marker + patients is shown in first column. Time-to-event distributions are exponential and all patients are followed to failure.
Randomize patient

Measure marker

Standard of care based rx (SOC-rx)

Marker based rx (M-rx)
Approximate number of events required for 80% power with 5% two-sided log-rank test for comparing arms of design shown in Figure 1. Randomized arms are mixtures of marker – and marker + patients. Hazard ratio for marker – patients is 1 for the two treatment groups and 0.67 for marker + patients. All patients are followed to failure.

<table>
<thead>
<tr>
<th>Proportion of Patients Marker +</th>
<th>Approximate Number of Events Required</th>
</tr>
</thead>
<tbody>
<tr>
<td>20%</td>
<td>5200</td>
</tr>
<tr>
<td>33%</td>
<td>1878</td>
</tr>
<tr>
<td>50%</td>
<td>820</td>
</tr>
</tbody>
</table>
Validation Study
Node negative Breast Cancer

- Prospective study design
- Samples collected and assayed from patients with node negative ER+ breast cancer receiving TAM
- Identify patients predicted to be very good prognosis on TAM alone using the single, fully specified multi-gene predictor of outcome
- Were long-term outcomes for patients in good prognosis group sufficiently good to have warranted withholding chemotherapy?
Validation Study
Node negative Breast Cancer

• Prospective study plan for use of archived specimens in a prospective clinical trial
• Samples collected and archived from patients who received Tam alone in prospective clinical trial
• Identify patients predicted to be very good prognosis on TAM alone using the single, fully specified multi-gene predictor of outcome developed externally to the trial
• Were long-term outcomes for patients in good prognosis group sufficiently good to have warranted withholding chemotherapy?
Assay Limitations of DNA Microarray Expression Profiling

- Need for fresh/frozen tumor
- Expression influenced by sample handling
- Assay variation among times and laboratories
• Some of the sources of assay variability will be controlled within a study but will limit the ability to accurately classify samples collected outside of study conditions
Validation Study for Identifying Node Positive Patients Who Benefit from a Specific Regimen

- Standard treatment C
- New treatment E
- Predictor based on previous data for identifying patients who benefit from E but not C
- Randomized study of E vs C
- Measure markers on all patients
- Compare E vs C separately within groups predicted to benefit from E and those not predicted to benefit from E
- Two clinical trials worth of patients
Conclusions

• New technology and biological knowledge make it increasingly feasible to identify which patients are most likely to benefit from a specified treatment

• “Predictive medicine” is feasible but does not mean “personalized treatment”

• Targeting treatment can greatly improve the therapeutic ratio of benefit to adverse effects
  – Smaller clinical trials needed
  – Treated patients benefit
  – Economic benefit for society
Conclusions

• Achieving the potential of new technology requires paradigm changes in focus and methods of “correlative science.”

• Achieving the potential of new technology requires paradigm changes in partnerships among industry, academia, and government.

• Effective interdisciplinary research requires increased emphasis on cross education of laboratory, clinical and statistical scientists.
Conclusions

• Prospectively specified analysis plans for phase III data are essential to achieve reliable results
  – Biomarker analysis does not mean exploratory analysis except in developmental studies
  – Biomarker classifiers used in phase III evaluations should be completely specified based on previous developmental studies


